

Amplite® Fluorimetric Glutathione Peroxidase Assay Kit *Blue Fluorescence*

Catalog number: 11560

Unit size: 200 Tests

Component	Storage	Amount (Cat No. 11560)
Component A: Glutathione Peroxidase Standard	Freeze (< -15 °C), Minimize light exposure	1 vial (0.5 U/vial)
Component B: Assay Buffer	Freeze (< -15 °C)	1 bottle (10 mL)
Component C: Enzyme Mix	Freeze (< -15 °C), Minimize light exposure	2 bottles (lyophilized powder)
Component D: GSH	Freeze (< -15 °C), Minimize light exposure	1 vial (3 mg/vial)
Component E: GPx Substrate	Freeze (< -15 °C), Minimize light exposure	1 vial (11 µL/vial)
Component F: Quest Fluor™ NADP Probe	Freeze (< -15 °C), Minimize light exposure	1 bottle (5 mL)
Component G: NADP Assay Solution	Freeze (< -15 °C), Minimize light exposure	1 bottle (5 mL)
Component H: Enhancer Solution	Freeze (< -15 °C), Minimize light exposure	1 bottle (3.5 mL)

OVERVIEW

Glutathione peroxidase is an enzyme family with peroxidase activity to protect the organism from oxidative damage. Glutathione peroxidase plays an important role in reducing organic hydroperoxides such as lipid hydroperoxides to their corresponding alcohols, or reducing free hydrogen peroxide to water. Glutathione peroxidase guards against oxidative damage to cell membranes and other oxidant-sensitive sites in cells. The altered glutathione peroxidase levels correlate with lesions caused by many common and complex diseases. Glutathione peroxidase level is measured in biological samples as a potential indicator for the potential treatment of cancer, diabetes, neurodegenerative and cardiovascular diseases. AAT Bioquest's Fluorimetric Glutathione Peroxidase Assay Kit offers a sensitive fluorimetric assay for measuring glutathione peroxidase levels in biological samples. This assay is based on the oxidation of glutathione (GSH) to oxidized glutathione (GSSG) catalyzed by glutathione peroxidase. The generated GSSG is recycled to its reduced state GSH by glutathione reductase (GR) and NADPH to generate NADP⁺ that can be specifically monitored using Quest Fluor™ NADP Probe, our newly developed proprietary NADP sensor. The NADP sensor reacts only with NADP to generate a fluorescent product. The fluorescence signal can be measured with a fluorescence microplate reader and is directly proportional to the glutathione peroxidase activity. Compared to other commercial kits that measure the decrease in absorbance of NADPH at 340 nm, our Quest Fluor™ NADP Probe can be used to quantify NADP level directly. With this fluorimetric glutathione peroxidase assay, as low as 1.25 mU/mL glutathione peroxidase can be detected in a 155 µL reaction volume.

AT A GLANCE
Protocol Summary

1. Prepare GPx standards or test samples (50 µL)
2. Add GPx working solution (50 µL)
3. Incubate at room temperature for 30 min
4. Add 20 µL Quest Fluor™ NADP Probe
5. Add 20 µL NADP Assay Solution
6. Incubate at room temperature for 10 - 20 min
7. Add 15 µL Enhancer Solution
8. Incubate at room temperature for 30 - 60 min
9. Record Fluorescence at Ex/Em= 420/480nm (Cutoff = 430 nm)

Important Note

To achieve the best results, it's strongly recommended to use the black plates. Thaw one vial of each kit component at room

temperature before starting the experiment.

KEY PARAMETERS
Fluorescence microplate reader

Cutoff	430 nm
Emission	480 nm
Excitation	420 nm
Recommended plate	Solid black

PREPARATION OF STOCK SOLUTIONS

Unless otherwise noted, all unused stock solutions should be divided into single-use aliquots and stored at -20 °C after preparation. Avoid repeated freeze-thaw cycles

Glutathione Peroxidase (GPx) standard solution (10 U/mL)

Add 50 µL of ddH₂O or 1× PBS buffer into the vial of GPx standard (Component A) to make 10 U/mL standard solution.

GSH stock solution (100X)

Add 100 µL of ddH₂O into the vial of GSH (Component D) to make 100X GSH stock solution.

GPx Substrate stock solution (100X)

Add 100 µL of ddH₂O into the vial of substrate (Component E) to make 100X GPx Substrate stock solution.

PREPARATION OF STANDARD SOLUTIONS

For convenience, use the Serial Dilution Planner:
<https://www.aatbio.com/tools/serial-dilution/11560>

GPx standard

Add 10 µL of GPx standard solution (10 U/mL) into 990 µL 1x PBS buffer + 0.1% BSA to generate 100 mU/mL GPx standard solution (GP7). Take 100 mU/mL GPx standard solution (GP7) and perform 1:1.5 serial dilutions in PBS + 0.1% BSA to get serial dilutions of GPx standard (GP6 - GP1). Note: Diluted GPx standard solution is unstable, and should be used within 4 hours.

PREPARATION OF WORKING SOLUTION

1. Add 5 mL of Assay Buffer (Component B) into a bottle of Enzyme Mix (Component C).
2. Add 50 µL GSH stock solution (Component D), 50 µL GPx

Substrate stock solution (Component E) into the bottle of Component B+C, and mix well to make GPx working solution (Component B+C+D+E). *Note:* This GPx working solution is enough for one 96-well plate. It is not stable; please use it promptly. It is not recommend storing unused GPx working solution.

EXAMPLE DATA ANALYSIS AND FIGURES

Placeholder for image details

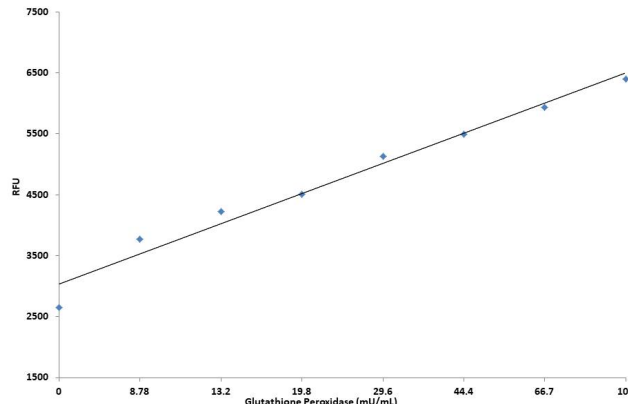


Figure 1. Glutathione Peroxidase (GPx) dose response was measured with Amplite® Fluorimetric Glutathione Peroxidase Assay Kit (Cat#11560) on a solid black 96-well plate using a Gemini microplate reader (Molecular Devices).

DISCLAIMER

AAT Bioquest provides high-quality reagents and materials for research use only. For proper handling of potentially hazardous chemicals, please consult the Safety Data Sheet (SDS) provided for the product. Chemical analysis and/or reverse engineering of any kit or its components is strictly prohibited without written permission from AAT Bioquest. Please call 408-733-1055 or email info@aatbio.com if you have any questions.

SAMPLE EXPERIMENTAL PROTOCOL

Table 1. Layout of GPx standards and test samples in a solid black 96-well microplate. GP= GPx Standards (GP1 - GP7, 8.78 to 100 mU/mL), BL=Blank Control, TS=Test Samples.

BL	BL	TS	TS
GP1	GP1
GP2	GP2
GP3	GP3		
GP4	GP4		
GP5	GP5		
GP6	GP6		
GP7	GP7		

Table 2. Reagent composition for each well.

Well	Volume	Reagent
GP1 - GP7	50 µL	Serial Dilution (8.78 to 100 mU/mL)
BL	50 µL	1X PBS Buffer
TS	50 µL	Test Sample

Run GPx assay:

1. Prepare GPx standards (GP), blank controls (BL), and test samples (TS) according to the layout provided in Tables 1 and 2. For a 384-well plate, use 25 µL of reagent per well instead of 50 µL.
2. Add 50 µL of GPx working solution to each well of GPx standard, blank control, and test samples to make the total assay volume of 100 µL/well. For a 384-well plate, add 25 µL GPx working solution into each well instead, for total of 50 µL/well.
3. Incubate the reaction at room temperature for 30 minutes, protected from light.

Run NADP assay:

1. Add 20 µL Quest Fluor™ NADP Probe (Component F) into each well of GPx standard, blank control, and test samples, mix well.
2. Add 20 µL NADP Assay Solution (Component G) into each well, mix well. For a 384-well plate, add 25 µL of sample and 10 µL of Quest Fluor™ NADP Probe (Component F) and 10 µL NADP Assay Solution (Component G) into each well.
3. Incubate the reaction at room temperature for 10 - 20 minutes, protected from light.
4. Add 15 µL Enhancer (Component H) to each well to make the total assay volume of 155 µL/well, and incubate at room temperature for 30 - 60 minutes, protected from light. For a 384-well plate, add 7.5 µL Enhancer.
5. Monitor the fluorescence increase with a fluorescence plate reader at Ex/Em = 420/480 nm (Cutoff = 430nm).