

**Fura-8FF™, AM**

 Catalog number: 20620  
 Unit size: 10x50 ug

Component	Storage	Amount (Cat No. 20620)
Fura-8FF™, AM	Freeze (< -15 °C), Minimize light exposure	10x50 ug

**OVERVIEW**

The cell-permeant Fura-8FF AM is an analog of Fura-8 AM with much lower calcium binding affinity,  $K_d \sim 10 \mu\text{M}$ . Fura-8FF has its emission shifted into longer visible wavelength that is compatible with the common filter sets. Fura-8FF™ AM is more sensitive to calcium than Fura-2FF AM with higher signal/background ratio than that of Fura-2FF AM. e., calculating the excitation intensity ratios at 354 nm and 415 nm by monitoring emission intensity at 530 nm.

**KEY PARAMETERS**
**Fluorescence microscope**

Emission	Fura 2 filter set
Excitation	Fura 2 filter set
Recommended plate	Black wall/clear bottom

**Fluorescence microplate reader**

Cutoff	475
Emission	530
Excitation	355, 415
Recommended plate	Black wall/clear bottom
Instrument specification(s)	Bottom read mode/Programmable liquid handling

**PREPARATION OF STOCK SOLUTIONS**

*Unless otherwise noted, all unused stock solutions should be divided into single-use aliquots and stored at -20 °C after preparation. Avoid repeated freeze-thaw cycles*

**Fura-8FF™ AM Stock Solution**

1. Prepare a 2 to 5 mM stock solution of Fura-8FF™ AM in high-quality, anhydrous DMSO.

**PREPARATION OF WORKING SOLUTION**
**Fura-8FF™ AM Working Solution**

1. On the day of the experiment, either dissolve Fura-8FF™ AM in DMSO or thaw an aliquot of the indicator stock solution to room temperature.
2. Prepare a 2 to 20  $\mu\text{M}$  Fura-8FF™ AM working solution in a buffer of your choice (e.g., Hanks and Hepes buffer) with 0.04% Pluronic® F-127. For most cell lines, Fura-8FF™ AM at a final concentration of 4-5  $\mu\text{M}$  is recommended. The exact concentration of indicators required for cell loading must be determined empirically.

**Note:** The nonionic detergent Pluronic® F-127 is sometimes used to increase the aqueous solubility of Fura-8FF™ AM. A variety of [Pluronic® F-127 solutions](#) can be purchased from AAT Bioquest.

**Note:** If your cells contain organic anion-transporters, probenecid (1-2 mM) may be added to the dye working solution (final in well concentration will be 0.5-1 mM) to reduce leakage of the de-esterified indicators. A variety of [ReadiUse™ Probenecid products](#), including water-soluble, sodium salt, and stabilized solutions, can be purchased from AAT Bioquest.

**SAMPLE EXPERIMENTAL PROTOCOL**

Following is our recommended protocol for loading AM esters into live cells. This protocol only provides a guideline and should be modified according to your specific needs.

1. Prepare cells in growth medium overnight.
2. On the next day, add 1X Fura-8FF™ AM working solution to your cell plate.

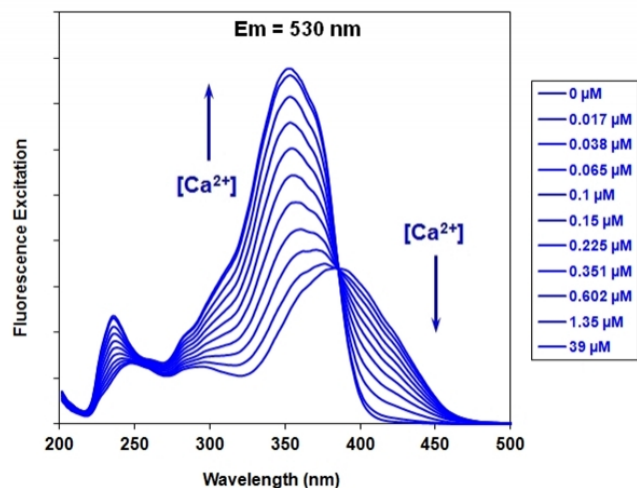
**Note:** If your compound(s) interfere with the serum, replace the growth medium with fresh HHBS buffer before dye-loading.

3. Incubate the dye-loaded plate in a cell incubator at 37 °C for 30 to 60 minutes.

**Note:** Incubating the dye for longer than 1 hour can improve signal intensities in certain cell lines.

4. Replace the dye working solution with HHBS or buffer of your choice (containing an anion transporter inhibitor, such as 1 mM probenecid, if applicable) to remove any excess probes.
5. Add the stimulant as desired and simultaneously measure fluorescence using either a fluorescence microscope equipped with a Fura 2 filter set or a fluorescence plate reader containing a programmable liquid handling system such as a FlexStation, at  $\text{Ex/Em}_1 = 355/530 \text{ nm}$  cutoff 475 nm and  $\text{Ex/Em}_2 = 415/530 \text{ nm}$  cutoff 475 nm.

## EXAMPLE DATA ANALYSIS AND FIGURES



**Figure 1.** Fluorescence excitation spectra of Fura-8™ in the presence of 0 to 39 μM free Ca<sup>2+</sup>.

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