

**MMP-3 Green™ substrate solution**

 Catalog number: 13527  
 Unit size: 100 Tests

| Component                       | Storage                                    | Amount    |
|---------------------------------|--|-----------|
| MMP-3 Green™ substrate Solution | Freeze (< -15 °C), Minimize light exposure | 100 tests |

**AT A GLANCE**
**Protocol Summary**

1. Add appropriate controls, or test samples (50 µL)
2. Pre-incubate for 10 - 15 minutes
3. Add MMP-3 Green™ substrate working solution (50 µL)
4. Skip incubation for kinetic reading or incubate 30 to 60 minutes for end point reading
5. Monitor fluorescence intensity at Ex/Em = 490/525 nm

**Important**

Thaw the solution at room temperature before starting the experiment. Prepare MMP-3 containing biological samples as desired.

**KEY PARAMETERS**
**Fluorescence microplate reader**

|                   |             |
|-------------------|-------------|
| Excitation        | 490 nm      |
| Emission          | 525 nm      |
| Cutoff            | 515 nm      |
| Recommended plate | Solid black |

**PREPARATION OF WORKING SOLUTION**
**1. MMP-3 Green™ Substrate working solution**

Add 50 µL of MMP-3 Green™ Substrate Solution into 5 mL of buffer of your choice to make a total volume of 5.05 mL. **Note:** Tris buffer can be used for the assay.

**2. MMP-3 dilutions**

Dilute MMP-3 to an appropriate concentration in buffer of your choice if purified MMP-3 is used. **Note:** MMP-3 needs to be activated before use. Avoid vigorous vortexing of the enzyme.

**3. Inhibitors and compounds dilution**

Make an appropriate concentration of known MMP-3 inhibitors and test compounds dilutions as desired if screening MMP-3 inhibitors.

**SAMPLE EXPERIMENTAL PROTOCOL**

1. Prepare MMP-3 containing biological samples as desired.
2. Activate pro-MMP-3 as per protocol. **Note:** Incubate the MMP-3 containing-samples or purified MMP-3 with equal volume of 2 mM APMA working solution (2X) at 37 °C for 24 hours. Activate MMP-3 immediately before the experiment.
3. Prepare controls and test samples (TS) according to the layout provided in Tables 1 and 2. For a 384-well plate, use 20 µL of reagent per well instead of 50 µL.
4. Pre-incubate the plate at a desired temperature for the enzyme reaction (e.g. 25 °C or 37 °C) for 10 - 15 minutes if you are screening MMP-3 inhibitors.
5. Add 50 µL (96-well) or 20 µL (384-well) of MMP-3 Green™ substrate working solution to the sample and control wells of the assay plate.

Mix the reagents well.

6. Monitor the fluorescence intensity with a fluorescence plate reader at Ex/Em = 490/525 nm. **For kinetic reading:** Immediately start measuring fluorescence intensity and continuously record data every 5 minutes for 30 to 60 minutes. **For end-point reading:** Incubate the reaction at room temperature for 30 to 60 minutes, kept from light if possible. Mix the reagents well, and then measure the fluorescence intensity. **Table 1.** Layout of the appropriate controls (as desired) and test samples in a 96-well microplate. SC= Substrate Control, IC= Inhibitor Control, VC=Vehicle Control, TC= Test Compound Control, TS=Test Samples.

|     |     |     |     |
|-----|-----|-----|-----|
| SC  | SC  | ... | ... |
| IC  | IC  |     |     |
| VC  | VC  |     |     |
| TC  | TC  |     |     |
| TS  | TS  |     |     |
| ... | ... |     |     |
| ... | ... |     |     |

**Table 2.** Reagent composition for each well.

| Well | Volume | Reagent  |
|------|--------|--|
| SC   | 50 µL  | Buffer of your choice                                    |
| IC   | 50 µL  | MMP-3 dilution and known MMP-3 inhibitor                 |
| VC   | 50 µL  | MMP-3 dilution and vehicle used to deliver test compound |
| TC   | 50 µL  | MMP-3 containing buffer and test compound                |
| TS   | 50 µL  | MMP-3 dilution with test compound                        |

**DISCLAIMER**

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