

Cell Meter™ Intracellular Fluorimetric Hydrogen Peroxide Assay Kit

Green Fluorescence

Ordering Information	Storage Conditions	Instrument Platform
Product Number: 11503 (200 assays)	Keep in freezer Avoid exposure to light	Fluorescence Microscope Fluorescence microplate readers

Introduction

Hydrogen peroxide (H₂O₂) is a reactive oxygen metabolic by-product that serves as a key regulator for a number of oxidative stress-related states. It is involved in many biological events that are linked to asthma, atherosclerosis, diabetic vasculopathy, osteoporosis, a number of neurodegenerative diseases and Down's syndrome. The measurement of this reactive species is helpful for determining how oxidative stress modulates various intracellular pathways.

This Cell Meter™ Intracellular Fluorimetric Hydrogen Peroxide Assay Kit uses our unique OxiVision™ Green Peroxide Sensor to quantify hydrogen peroxide in live cells. OxiVision™ Green peroxide sensor is cell-permeable, and generates the green fluorescence when it reacts with hydrogen peroxide. This kit provides a sensitive tool to monitor hydrogen peroxide level in living cells. The kit is also optimized with “mix and read” assay format for solution based assay. It provides a sensitive, one-step fluorimetric assay to detect as little as 0.3 nanomoles of H₂O₂ in a 100 µL assay volume (3 µM). The assay can be performed in a convenient 96-well or 384-well microtiter-plate format. Its signal can be easily read by either a fluorescence microplate reader at Ex/Em = 490/520 nm for H₂O₂ detection in solution or a fluorescence microscopy and a flow cytometry for live cell H₂O₂ detection.

Kit Key Features

Broad Application:	Can be used for quantifying hydrogen peroxide in live cells, in solutions, and in cell extracts.
Continuous:	Easily adapted to automation without a separation step.
Convenient:	Formulated to have minimal hands-on time. No wash is required.

Kit Components

Components	Amount
Component A: OxiVision™ Green Peroxide Sensor	1 vial
Component B: H ₂ O ₂	1 vial (3% stabilized solution, 200 µL)
Component C: Assay Buffer	1 bottle (20 mL)
Component D: DMSO	1 vial (200 µL)

Assay Protocol for One 96-Well Plate

Brief Summary for Solution Assay

Prepare H₂O₂ reaction mixture (50 µL) → Add H₂O₂ standards or test samples (50 µL) → Incubate at room temperature for 15-60 minutes → Read fluorescence intensity at Ex/Em = 490/520 nm

Note: Thaw all the kit components at room temperature before starting the experiment.

1. Prepare stock solutions:

- 1.1 OxiVision™ Green Peroxide Sensor stock solution (250X): Add 50 µL of DMSO (Component D) into the vial of OxiVision™ Green Peroxide Sensor (Component A). The stock solution should be used promptly. Any remaining solution should be aliquoted and refrozen at -20 °C.

Note: Avoid repeated freeze-thaw cycles and protect from light.

- 1.2 20 mM H₂O₂ stock solution: Add 22.7 µL of 3% H₂O₂ (0.88 M, Component B) into 977 µL of Assay Buffer (Component C).

Note: The diluted H₂O₂ solution is not stable. The unused portion should be discarded.

2. Prepare 1X OxiVision™ Green Peroxide Sensor working solution:

Add 20 µL of OxiVision™ Green Peroxide Sensor stock solution (250X, from Step 1.1) into 5 mL of Assay Buffer (Component C).

3. Prepare serially diluted H₂O₂ standards (0 to 1000 µM):

- 3.1 Add 50 µL of 20 mM H₂O₂ solution (from Step 1.2) into 950 µL of Assay Buffer (Component C) to get 1000 µM H₂O₂ solution.
- 3.2 Take 200 µL of 1000 µM H₂O₂ solution to perform 1:3 serial dilutions to get 300, 100, 30, 10, 3, 1, 0.3 and 0 µM serially diluted H₂O₂ stands.
- 3.3 Add H₂O₂ standards and H₂O₂-containing test samples into a solid black 96-well microplate as described in Tables 1 and 2.

Table 1. Layout of H₂O₂ standards and test samples in a solid black 96-well microplate

BL	BL	TS	TS						
HS1	HS1						
HS2	HS2										
HS3	HS3										
HS4	HS4										
HS5	HS5										
HS6	HS6										
HS7	HS7										

Note: HS= H₂O₂ Standards; BL=Blank Control; TS=Test Samples

Table 2. Reagent composition for each well

H ₂ O ₂ Standards	Blank Control	Test Sample
Serial Dilutions*: 50 µL	Assay Buffer (Component C): 50 µL	50 µL

4. Run H₂O₂ assay in supernatants reaction:

- 4.1 Add 50 µL of 1X OxiVision™ Green Peroxide Sensor working solution (from Step 2) to each well of the H₂O₂ standard, blank control, and test samples (see Step 3.3) to make the total H₂O₂ assay volume of 100 µL/well.

Note: For a 384-well plate, add 25 µL of sample and 25 µL of 1X OxiVision™ Green peroxide Sensor working solution into each well.

- 4.2 Incubate the reaction at room temperature for 15 to 30 minutes, protected from light.
- 4.3 Monitor the fluorescence increase at Ex/Em = 490±10/520±10 nm (optimal Ex/Em = 490/520) with a fluorescence plate reader.

5. Run H₂O₂ assay in live cells:**Brief Summary for Live Cell Assay**

Prepare cells in growth medium → Stain cells with OxiVision™ Green Peroxide Sensor → Treat cells with test compounds → Monitor fluorescence intensity at Ex/Em = 490/520 nm

OxiVision™ Green Peroxide Sensor can be loaded passively into living cells and report the micromolar changes in intracellular H₂O₂ concentrations. The following is a suggested microscope imaging protocol that can be modified to meet specific research needs.

- 5.1 Treat the cells as desired.
- 5.2 Wash the cells with PBS buffer, incubated the cells with 100 µL/well 1X OxiVision™ Green Peroxide Sensor working solution (from Step 2) for 5 to 60 minutes or your desired time.
Note: For a 384-well plate, add 25 µL/well of 1X OxiVision™ Green Peroxide Sensor working solution.
- 5.3 Monitor the fluorescence increase at excitation 490 nm and emission at 525nm using a fluorescence plate reader with bottom read mode. Or image the fluorescence change with a fluorescence microscope using FITC channel.

Data Analysis

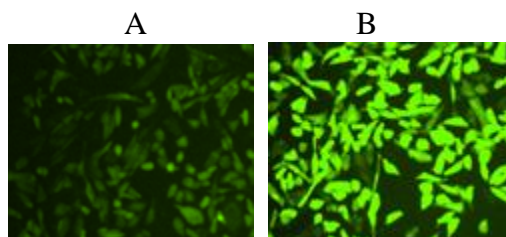


Figure 1. Fluorescence images of Live CHO-K1 cells in a Costar black 96-well plate. Live CHO-K1 cells were stained with Cell Meter™ Intracellular Fluorimetric Hydrogen Peroxide Assay Kit. A: Control cells. B: Cells treated with 100 μM H_2O_2 at room temperature for 5 minutes.

References

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5. Ma HP. [Hydrogen peroxide stimulates the epithelial sodium channel through a phosphatidylinositol 3-kinase-dependent pathway](#) *J Biol Chem.* 2011, 286(37):32444-53. doi: 10.1074/jbc.M111.254102.

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