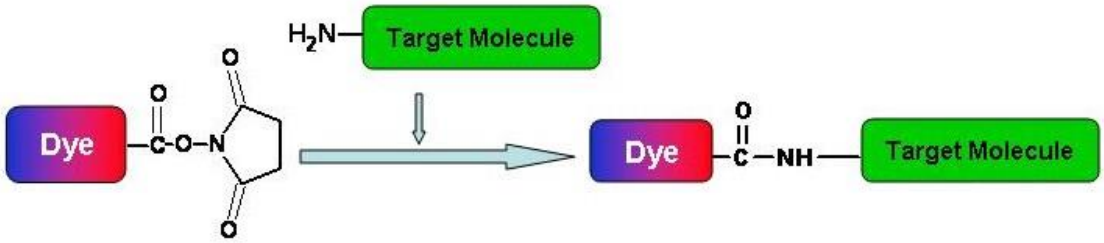
Labeling Proteins with Protonex™ Dye SE

Introduction

Protonex™ dyes conjugates demonstrated pH-dependent fluorescence. Unlike most of the existing fluorescent dyes that are more fluorescent at higher pH, acidic conditions enhance the fluorescence of Protonex™ dye conjugates. The fluorescence of Protonex™ dye conjugates dramatically increases as pH decreases from neutral to the acidic, making them a robust set of tools to study phagocytosis and its regulation by drugs and/or environmental factors. The lack of fluorescence outside the cells eliminates the wash steps. Protonex™ dye conjugates provide a set of powerful tools to study phagocytosis and to monitor acidic cell compartments such as endosomes and lysosomes. Protonex™ dye conjugates are non-fluorescent outside the cells, but fluoresce brightly in acidic compartments (such as phagosomes, lysosomes and endosomes). Protonex™ SE dyes can be readily used to make a variety of bioconjugates for imaging or flow applications, enabling the specific detection of phagocytosis and endocytosis with reduced signal variability and improved accuracy. These conjugates can be also used for multiplexing cell functional analysis. Protonex™ Red has the spectral properties similar to those of Texas Red, and Protonex™ Green has the spectral properties similar to those of FITC making the common filter sets of Texas Red and FITC readily available to the assays of Protonex™ Red and Protonex™ Green respectively.

Succinimidyl (NHS) esters are proven to be the best reagents for amine modifications because the amide bonds that are formed are essentially identical to, and as stable as the natural peptide bonds. These reagents are generally stable and show good reactivity and selectivity with aliphatic amines. There are few factors that need be considered when succinimidyl esters compounds are used for conjugation reaction: 1). *Solvents:* For the most part, reactive dyes should be dissolved in anhydrous dimethylformamide (DMF) or dimethylsulfoxide (DMSO). 2). *Reaction pH:* The labeling reactions of amines with succinimidyl esters are strongly pH dependent. Amine-reactive reagents react with non-protonated aliphatic amine groups, including the terminal amines of proteins and the ε-amino groups of lysines. Thus amine acylation reactions are usually carried out above pH 7.5. Protein modifications by succinimidyl esters can typically be done at pH 8.5-9.5. 3). *Reaction Buffers:* Buffers that contain free amines such as Tris and glycine and thiol compounds must be avoided when using an amine-reactive reagent. Ammonium salts (such as ammonium sulfate and ammonium acetate) that are widely used for protein precipitation must also be removed (such as via dialysis) before performing dye conjugations. 4). *Reaction Temperature:* Most conjugations are done at room temperature. However, either elevated or reduced temperature may be required for a particular labeling reaction.



Storage and Handling

Upon receipt, Protonex™ dyes should be stored at <-15 °C, and kept from light and moisture. The reconstituted DMSO stock solution of Protonex ™ dyes can be stored at <-15 °C for less than two weeks. The protein conjugate should be stored at > 0.5 mg/mL in the presence of a carrier protein (e.g., 0.1% bovine serum albumin). The conjugate solution could be stored at 4 °C for two months without significant change when stored in the presence of 2 mM sodium azide and kept from light. For longer storage, the protein conjugates could be lyophilized or divided into single-used aliquots and stored at ≤ –60 °C, and protected from light.

Chemical and Physical Properties (Table 1)

Cat#	Protonex™ Dyes	MW	Solvent	Ex (nm)	Em (nm)	Abs (nm)	Extinction Coefficient[1]	CF at 260 nm[2]	CF at 280 nm[3]
21208	Protonex™ Red 600, SE	953.06	DMSO	575	597	575	95,000	0.297	0.213
21216	Protonex™ Green 500, SE	539.54	DMSO	443	505	457	20,000	1.487	1.069

Note 1. *Extinction coefficient at their maximum absorption wavelength; 2. CF at 260 nm is the correction factor used for eliminating the dye contribution to the absorbance at 260 nm (for oligo and nucleic acid labeling); 3. CF at 280 nm is the correction factor used for eliminating the dye contribution to the absorbance at 280 nm (for peptide and protein labeling).*

Note 2. *The buffer for the absorbance and extinction coefficient measurements is 100 mM MES buffer (pH 4.7).*

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Sample Labeling Protocol

Note: This labeling protocol was developed for the conjugate of Goat anti-mouse IgG with Protonex™ Green 500, SE. You might need further optimization for your particular proteins.

1. Prepare protein stock solution (Solution A):

Mix 100 µL of a reaction buffer (e.g., 1 M sodium carbonate solution or 1 M phosphate buffer with pH ~9.0) with 900 µL of the target protein solution (e.g. antibody, protein concentration >2 mg/ml if possible) to give 1 mL protein labeling stock solution.

Note 1*: The pH of the protein solution (Solution A) should be 8.5 ± 0.5. If the pH of the protein solution is lower than 8.0, adjust the pH to the range of 8.0-9.0 using 1 M sodium bicarbonate solution or 1 M pH 9.0 phosphate buffer.*
Note 2*: The protein should be dissolved in 1X phosphate buffered saline (PBS), pH 7.2-7.4. If the protein is dissolved in Tris or glycine buffer, it must be dialyzed against 1X PBS, pH 7.2-7.4, to remove free amines or ammonium salts (such as ammonium sulfate and ammonium acetate) that are widely used for protein precipitation.*
Note 3*: Impure antibodies or antibodies stabilized with bovine serum albumin (BSA) or gelatin will not be labeled well. The presence of sodium azide or thimerosal might also interfere with the conjugation reaction. Sodium azide or thimerosal can be removed by dialysis or spin column for optimal labeling results.*
Note 4*: The conjugation efficiency is significantly reduced if the protein concentration is less than 2 mg/mL. For optimal labeling efficiency the final protein concentration range of 2-10 mg/mL is recommended.*

2. Prepare dye stock solution (Solution B):

Add anhydrous DMSO into the vial of Protonex™ dye SE to make a 10-20 mM stock solution. Mix well by pipetting or vortex.

Note 1: *Prepare the dye stock solution (Solution B) before starting the conjugation. Use promptly. Extended storage of the dye stock solution may reduce the dye activity. Solution B can be stored in freezer for two weeks when kept from light and moisture. Avoid freeze-thaw cycles.*

Note 2: *The Protonex™ Green 500 SE is not very soluble in DMSO, it is OK to use its suspension in the conjugation buffer.*

3. Determine the optimal dye/protein ratio (optional):

Note: *Each protein requires distinct dye/protein ratio, which also depends on the properties of dyes. Over labeling of a protein could detrimentally affects its binding affinity while the protein conjugates of low dye/protein ratio gives reduced sensitivity. We recommend you experimentally determine the best dye/protein ratio by repeating Steps 4 and 5 using a serial different amount of labeling dye solutions. In general 4-6 dyes/protein are recommended for most of dye-protein conjugates.*

3.1 Use 10:1 molar ratio of Solution B (dye)/Solution A (protein) as the starting point: Add 5 µl of the dye stock solution (Solution B, assuming the dye stock solution is 10 mM) into the vial of the protein solution (95 µl of Solution A) with effective shaking. The concentration of the protein is ~0.05 mM assuming the protein concentration is 10 mg/mL and the molecular weight of the protein is ~200KD.

Note: *The concentration of the DMSO in the protein solution should be <10 %.*

3.2 Run conjugation reaction (see Step 4 below).

3.3 Repeat #3.2 with the molar ratios of Solution B/Solution A at 5:1; 15:1 and 20:1 respectively.

3.4 Purify the desired conjugates using premade spin columns.

3.5 Calculate the dye/protein ratio (DOS) for the above 4 conjugates (see next page).

3.6 Run your functional tests of the above 4 conjugates to determine the best dye/protein ratio to scale up your labeling reaction.

4. Run conjugation reaction:

4.1 Add the appropriate amount of dye stock solution (Solution B) into the vial of the protein solution (Solution A) with effective shaking.

Note: *The best molar ratio of Solution B/Solution is determined from Step 3.6. If Step 3 is skipped, we recommend to use 10:1 molar ratio of Solution B (dye)/Solution A (protein).*

4.2 Continue to rotate or shake the reaction mixture at room temperature for 30-60 minutes.

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5. Purify the conjugation

The following protocol is an example of dye-protein conjugate purification by using a Sephadex G-25 column.

5.1 Prepare Sephadex G-25 column according to the manufacture instruction.

5.2 Load the reaction mixture (directly from Step 4) to the top of the Sephadex G-25 column.

5.3 Add PBS (pH 7.2-7.4) as soon as the sample runs just below the top resin surface.

5.4 Add more PBS (pH 7.2-7.4) to the desired sample to complete the column purification. Combine the fractions that contain the desired dye-protein conjugate.

Note 1: *For immediate use, the dye-protein conjugate need be diluted with staining buffer, and aliquoted for multiple uses.*
Note 2: *For longer term storage, dye-protein conjugate solution need be concentrated or freeze dried (see below).*

Characterize the Desired Dye-Protein Conjugate

The Degree of Substitution (DOS) is the most important factor for characterizing dye-labeled protein. Proteins of lower DOS usually have weaker fluorescence intensity, but proteins of higher DOS (e.g. DOS > 6) tend to have reduced fluorescence too. The optimal DOS for most antibodies is recommended between 2 and 10 depending on the properties of dye and protein. For effective labeling, the degree of substitution should be controlled to have 4-10 moles of iFluor™ 647 SE to one mole of antibody. The following steps are used to determine the DOS of iFluor™ 647 SE labeled proteins.

1. Measure absorption:

To measure the absorption spectrum of a dye-protein conjugate, it is recommended to keep the sample concentration in the range of 1-10 μM depending on the extinction coefficient of the dye.

2. Read OD (absorbance) at 280 nm and dye maximum absorption:

Measure the absorbance (OD) of the Protonex™ dyes conjugates at 280 nm (the maximum absorption of protein) and at the λ_{max} for the Protonex™ dyes. For most spectrophotometers, the sample (from the column fractions) need be diluted with de-ionized water* so that the OD values are in the range of 0.1 to 0.9. To obtain accurate DOS, make sure that the conjugate is free of the non-conjugated dye. See the absorbance maxima, the extinction coefficient, and the correction factor for the pHrodo™ dyes on Table1.

3. Calculate DOS using the following equations:

3.1 Calculate protein concentration $[\text{Protein}] = \frac{\text{A280} - (\text{OD @ Dye Maximum Absoprtion X CF@280nm})}{\text{Protein Extinction Coefficient}} \times \text{dilution factor}$

3.2 Calculate dye concentration $[\text{Dye}] = \frac{\text{OD @ Maximum Absoprtion}}{\text{Dye Extinction Coefficient}} \times \text{dilution factor}$

3.3 Calculate the degree of labeling DOS = [Dye]/[Protein] $= [{}^{D}OD_{max} \times {}^{P}\varepsilon_{280}] / [\text{ Dye Extinction Coefficient} \times (A_{280} - \text{CF@280nm X } A_{max})]$

[Dye] is the dye concentration, and can be readily calculated from the Bee-Lambert Law: $A=\varepsilon_{dye}CL$. [Protein] is the protein concentration. This value can be either estimated by the weight (added to the reaction) if the conjugation efficiency is high enough (preferably > 70%) or more accurately calculated by the Beer-Lambert Law: $A=\varepsilon_{protein}CL$. For example, IgG has the ε value to be 203,000 $cm^{-1}M^{-1}$. ${}^{P}\varepsilon_{280}$ = protein molar extinction coefficient at 280 nm (e. g. the molar extinction coefficient of IgG is 203,000 $cm^{-1}M^{-1}$). CF (dye absorption correction factor at 280 nm) .

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