

Actin Lightning™ iFluor® 570 Phalloidin Conjugate

Catalog number: 22716
Unit size: 100 Tests

Component	Storage	Amount (Cat No. 22716)
Actin Lightning™ iFluor® 570 Phalloidin Conjugate	Freeze (< -15 °C), Minimize light exposure	100 Tests

OVERVIEW

Actin Lightning™ iFluor® 570 Phalloidin Conjugate is a novel fluorescent phalloidin conjugate designed for high-affinity labeling of filamentous actin (F-actin). This high-affinity design supports bright and specific F-actin staining, allowing stained samples to be imaged for around one month after staining with minimal loss of signal or specificity under appropriate mounting and storage conditions. This conjugate is labeled with iFluor® 570, an orange fluorophore with excitation and emission maxima of 557/570 nm. It is suitable for fluorescence microscopy workflows involving fixed cells and tissue sections, and provides an orange-channel option for imaging cell morphology, cytoskeletal organization, and F-actin distribution.

AT A GLANCE

Protocol Summary

1. Prepare samples in microplate wells
2. Remove liquid from samples in the plate
3. Add Actin Lightning™ iFluor® 570 Phalloidin Conjugate working solution (100 µL/well)
4. Stain the cells at room temperature for 20 to 90 minutes
5. Wash the cells and examine the specimen under microscope with Cy3 filter

Important Note

Warm the vial to room temperature and centrifuge briefly before opening.

Storage and Handling Conditions

The solution should be stable for at least 6 months if stored at -20 °C. Protect the fluorescent conjugates from light, and avoid freeze/thaw cycles.

Note: Phalloidin is toxic, although the amount of toxin present in a vial could be lethal only to a mosquito (LD50 of phalloidin = 2 mg/kg), it should be handled with care.

KEY PARAMETERS

Fluorescence microscope

Excitation	Cy3 filter set
Emission	Cy3 filter set
Recommended plate	Black wall/clear bottom

PREPARATION OF WORKING SOLUTION

Actin Lightning™ iFluor® 570 Phalloidin Conjugate Working Solution (1X):

Prepare working solution by adding 1 µL of Actin Lightning™ iFluor® 570 Phalloidin Conjugate stock solution into 1 mL of PBS with 1% BSA.

Note: Different cell types might be stained differently. The concentration of phalloidin conjugate working solution should be prepared accordingly.

SAMPLE EXPERIMENTAL PROTOCOL

This protocol only provides a guideline, and should be modified according to your specific needs.

Cell Staining

1. Prepare cells in growth medium overnight.
2. Remove the cell culture medium and wash cells with HHBS or buffer of your choice.
3. Perform formaldehyde fixation. Incubate cells with 3.0–4.0 % formaldehyde in PBS at room temperature for 10–30 minutes.

Note: Avoid any methanol containing fixatives since methanol can disrupt actin during the fixation process. The preferred fixative is methanol-free formaldehyde.

4. Rinse the fixed cells 2–3 times in PBS.
5. **Optional:** Add 0.1% Triton X-100 in PBS into fixed cells for 3 to 5 minutes to increase permeability. Rinse the cells 2–3 times in PBS.
6. Add 100 µL of Actin Lightning™ iFluor® 570 Phalloidin Conjugate working solution (1X).
7. Incubate cells in a cell incubator at room temperature for 20 to 90 minutes.
Note: Incubating the dye for longer than 60 minutes can improve signal intensities in certain cell lines.
8. Remove the dye working solution and wash cells with PBS to remove any excess probes.
9. Image cells with fluorescence microscope using Cy3 filter set.

EXAMPLE DATA ANALYSIS AND FIGURES

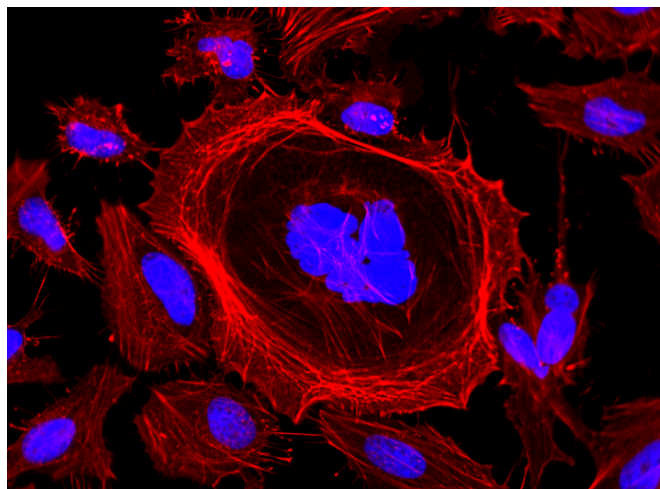


Figure 1. Fluorescence image of HeLa cells stained with Actin Lightning™ iFluor® 570 Phalloidin Conjugate and Nuclear Blue™ DCS1. Cells were fixed with 4% formaldehyde, permeabilized with 0.2% Triton™ X-100, then stained with Actin Lightning™ iFluor® 570

Phalloidin Conjugate (Cat. #22716) for F-actin detection and counterstained with Nuclear Blue™ DCS1 (Cat. #17548). Images were acquired using Cy3 and DAPI filter sets and overlaid to show F-actin in red and nuclei in blue.

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