

## Cal-630™ AM

Catalog number: 20530, 20531, 20532  
 Unit size: 5x50 ug, 10x50 ug, 1 mg

| Component   | Storage                                   | Amount (Cat No. 20530) | Amount (Cat No. 20531) | Amount (Cat No. 20532) |
|-------------|---|------------------------|------------------------|------------------------|
| Cal-630™ AM | Freeze (<-15 °C), Minimize light exposure | 5x50 ug                | 10x50 ug               | 1 vial (1 mg)          |

### OVERVIEW

Calcium measurement is critical for numerous biological investigations. Fluorescent probes that show spectral responses upon binding calcium have enabled researchers to investigate changes in intracellular free calcium concentrations by using fluorescence microscopy, flow cytometry, fluorescence spectroscopy and fluorescence microplate readers. x-Rhod-1 is commonly used as a red fluorescent calcium indicator. However, x-Rhod-1 is only moderately fluorescent in live cells upon esterase hydrolysis, and has very small cellular calcium responses. Cal-630™ has been developed to improve x-Rhod-1 cell loading and calcium response while maintaining the spectral wavelength of x-Rhod-1, making it compatible with Texas Red® filter set. In CHO and HEK cells Cal-630™ AM has cellular calcium response that is much more sensitive than x-Rhod-1. The spectra of Cal-630 is well separated from those of FITC, Alexa Fluor® 488 and GFP, making it an ideal calcium probe for multiplexing intracellular assays with GFP cell lines or FITC/Alexa Fluor® 488 labeled antibodies.

### KEY PARAMETERS

#### Fluorescence microscope

|                   |                         |
|-------------------|-------------------------|
| Emission          | Texas Red               |
| Excitation        | Texas Red               |
| Recommended plate | Black wall/clear bottom |

#### Fluorescence microplate reader

|                             |   |
|-----------------------------|---|
| Cutoff                      | 630   |
| Emission                    | 640   |
| Excitation                  | 600   |
| Recommended plate           | Black wall/clear bottom                       |
| Instrument specification(s) | Bottom read mode/Programmable liquid handling |

### PREPARATION OF STOCK SOLUTIONS

Unless otherwise noted, all unused stock solutions should be divided into single-use aliquots and stored at -20 °C after preparation. Avoid repeated freeze-thaw cycles

#### Cal-630™ AM Stock Solution

1. Prepare a 2 to 5 mM stock solution of Cal-630™ AM in anhydrous DMSO.

**Note:** When reconstituted in DMSO, Cal-630™ AM is a clear, colorless solution.

### PREPARATION OF WORKING SOLUTION

#### Cal-630™ AM Working Solution

1. On the day of the experiment, either dissolve Cal-630™ AM in DMSO or thaw an aliquot of the indicator stock solution to room temperature.
2. Prepare a 2 to 20 µM Cal-630™ AM working solution in a buffer of

your choice (e.g., Hanks and Hepes buffer) with 0.04% Pluronic® F-127. For most cell lines, Cal-630™ AM at a final concentration of 4-5 µM is recommended. The exact concentration of indicators required for cell loading must be determined empirically.

**Note:** The nonionic detergent Pluronic® F-127 is sometimes used to increase the aqueous solubility of Cal-630™ AM. A variety of Pluronic® F-127 solutions can be purchased from AAT Bioquest.

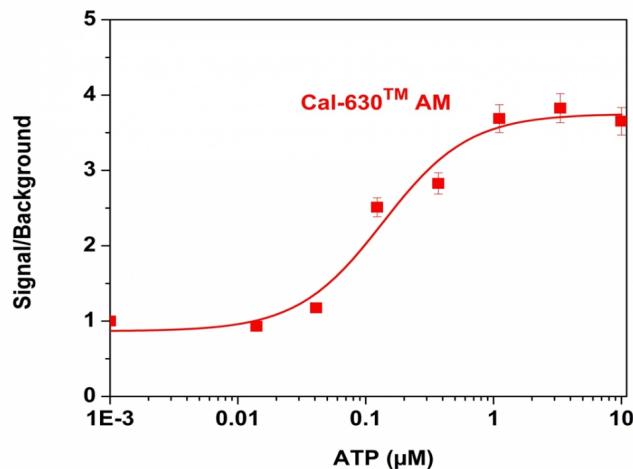
**Note:** If your cells contain organic anion-transporters, probenecid (1-2 mM) may be added to the dye working solution (final in well concentration will be 0.5-1 mM) to reduce leakage of the de-esterified indicators. A variety of [ReadiUse™ Probenecid products](#), including water-soluble, sodium salt, and stabilized solutions, can be purchased from AAT Bioquest.

### SAMPLE EXPERIMENTAL PROTOCOL

Following is our recommended protocol for loading AM esters into live cells. This protocol only provides a guideline and should be modified according to your specific needs.

1. Prepare cells in growth medium overnight.
2. On the next day, add 1X Cal-630™ AM working solution to your cell plate.
- Note:** If your compound(s) interfere with the serum, replace the growth medium with fresh HHBS buffer before dye-loading.
3. Incubate the dye-loaded plate in a cell incubator at 37 °C for 30 to 60 minutes.
- Note:** Incubating the dye for longer than 2 hours can improve signal intensities in certain cell lines.
4. Replace the dye working solution with HHBS or buffer of your choice (containing an anion transporter inhibitor, such as 1 mM probenecid, if applicable) to remove any excess probes.
5. Add the stimulant as desired and simultaneously measure fluorescence using either a fluorescence microscope equipped with a Texas Red filter set or a fluorescence plate reader containing a programmable liquid handling system such as an FDSS, FLIPR, or FlexStation, at Ex/Em = 600/640 nm cutoff 630 nm.

## EXAMPLE DATA ANALYSIS AND FIGURES



**Figure 1.** ATP-stimulated calcium responses of endogenous P2Y receptor in CHO-K1 cells incubated with Cal-630™ AM (red curve). CHO-K1 cells were seeded overnight at 50,000 cells per 100  $\mu$ L per well in a Costar black wall/clear bottom 96-well plate. 100  $\mu$ L of 5  $\mu$ M Cal-630™ AM in HHBS (with 1.0 mM probenecid) was added into the cells and incubated at 37 °C for 1 hour. ATP (50  $\mu$ L/well) was added using FlexStation to achieve the final indicated concentrations.

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