Fluo-5F, AM *Cell permeant*

Catalog number: 20560
Unit size: 10x50 ug

Component	Storage	Amount (Cat No. 20560)
Fluo-5F, AM *Cell permeant*	Freeze (< -15 °C), Minimize light exposure	10x50 ug

OVERVIEW

Fluo-5F is an analog of Fluo-4 with lower calcium-binding affinity (Kd = ~2.3 uM), making it suitable for detecting intracellular calcium levels in the range of 1 µM to 1 mM that would saturate the response of Fluo-4. Cells may be loaded with Fluo-5F AM ester by adding the dissolved indicator directly to dishes containing cultured cells. It is compatible with excitation at 488 nm by argon-ion laser sources, making Fluo-5F useful for confocal microscopy, flow cytometry, and microplate screening applications. It has excitation and emission wavelengths at 494 and 516 nm respectively. Upon calcium binding, its fluorescence intensity increases by >100 fold.

KEY PARAMETERS

Flow cytometer

Emission	530/30 nm filter
Excitation	488 nm laser
Instrument specification(s)	FITC channel

Fluorescence microscope

Emission	FITC
Excitation	FITC
Recommended plate	Black wall/clear bottom

Fluorescence microplate reader

Cutoff	515
Emission	525
Excitation	490
Recommended plate	Black wall/clear bottom
Instrument specification(s)	Bottom read mode/Programmable liquid handling

PREPARATION OF STOCK SOLUTIONS

Unless otherwise noted, all unused stock solutions should be divided into single-use aliquots and stored at -20 °C after preparation. Avoid repeated freeze-thaw cycles

Fluo-5F AM Stock Solution

1. Prepare a 2 to 5 mM stock solution of Fluo-5F AM in high-quality, anhydrous DMSO.

PREPARATION OF WORKING SOLUTION

Fluo-5F AM Working Solution

1. On the day of the experiment, either dissolve Fluo-5F AM in DMSO or thaw an aliquot of the indicator stock solution to room temperature.

2. Prepare a 2 to 20 µM Fluo-5F AM working solution in a buffer of your choice (e.g., Hanks and Hepes buffer) with 0.04% Pluronic® F-127. For most cell lines, Fluo-5F AM at a final concentration of 4-5 µM is adequate. The optimal concentration for cell loading must be determined empirically.

   Note: The nonionic detergent Pluronic® F-127 is sometimes used to increase the aqueous solubility of Fluo-5F AM. A variety of Pluronic® F-127 solutions can be purchased from AAT Bioquest.

   Note: If your cells contain organic anion-transporters, probenecid (1-2 mM) may be added to the dye working solution (final in well concentration will be 0.5-1 mM) to reduce leakage of the de-esterified indicators. A variety of ReadiUse™ Probenecid products, including water-soluble, sodium salt, and stabilized solutions, can be purchased from AAT Bioquest.

SAMPLE EXPERIMENTAL PROTOCOL

Following is our recommended protocol for loading AM esters into live cells. This protocol only provides a guideline and should be modified according to your specific needs.

1. Prepare cells in growth medium overnight.

2. On the next day, add 1X Fluo-5F AM working solution to your cell plate.

   Note: If your compound(s) interfere with the serum, replace the growth medium with fresh HHBS buffer before dye-loading.

3. Incubate the dye-loaded plate in a cell incubator at 37 °C for 30 to 60 minutes.

   Note: Incubating the dye for longer than 2 hours can improve signal intensities in certain cell lines.

4. Replace the dye working solution with HHBS or buffer of your choice (containing an anion transporter inhibitor, such as 1 mM probenecid, if applicable) to remove any excess probes.

5. Add the stimulant as desired and simultaneously measure fluorescence using either a fluorescence microscope equipped with a FITC filter set or a fluorescence plate reader containing a programmable liquid handling system such as an FDSS, FLIPR, or FlexStation, at 490/525 nm cutoff 515 nm.

EXAMPLE DATA ANALYSIS AND FIGURES

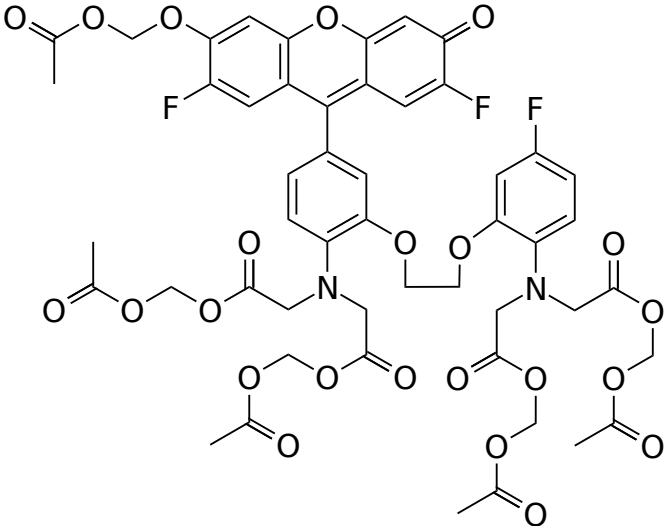

Figure 1. Chemical structure for Fluo-5F, AM *Cell permeant*

DISCLAIMER

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