

## Mag-Fluo-4 AM

Catalog number: 20401  
Unit size: 10x50 ug

Component	Storage	Amount (Cat No. 20401)
Mag-Fluo-4 AM	Freeze (< -15 °C), Minimize light exposure	10x50 ug

### OVERVIEW

The cell-permeant Mag-Fluo-4 AM is an analog of Fluo-4 AM with a Kd for Mg ion of 4.7 mM and a Kd for Ca ion of 22  $\mu$ M, making it useful as an intracellular Mg ion indicator as well as a low-affinity Ca ion indicator. This low-affinity fluorescent Ca ion indicator has been used to accurately track the kinetics of the spatially averaged free Ca ion transient in skeletal muscle. Mag-Fluo-4 yields reliable kinetic information about the spatially averaged free Ca ion transient in skeletal muscle.

to increase the aqueous solubility of Mag-Fluo-4 AM. A variety of Pluronic® F-127 solutions can be purchased from AAT Bioquest.

**Note:** If your cells contain organic anion-transporters, probenecid (1-2 mM) may be added to the dye working solution (final in well concentration will be 0.5-1 mM) to reduce leakage of the de-esterified indicators. A variety of [ReadiUse™ Probenecid products](#), including water-soluble, sodium salt, and stabilized solutions, can be purchased from AAT Bioquest.

### KEY PARAMETERS

#### Flow cytometer

Emission	530/30 nm filter
Excitation	488 nm laser
Instrument specification(s)	FITC channel

#### Fluorescence microscope

Emission	FITC filter set
Excitation	FITC filter set
Recommended plate	Black wall/clear bottom

#### Fluorescence microplate reader

Cutoff	515
Emission	525
Excitation	490
Recommended plate	Black wall/clear bottom
Instrument specification(s)	Bottom read mode/Programmable liquid handling

### SAMPLE EXPERIMENTAL PROTOCOL

Following is our recommended protocol for loading AM esters into live cells. This protocol only provides a guideline and should be modified according to your specific needs.

1. Prepare cells in growth medium overnight.
2. On the next day, add 1X Mag-Fluo-4 AM working solution to your cell plate.

**Note:** If your compound(s) interfere with the serum, replace the growth medium with fresh HHBS buffer before dye-loading.

3. Incubate the dye-loaded plate in a cell incubator at 37 °C for 30 to 60 minutes.

**Note:** Incubating the dye for longer than 1 hour can improve signal intensities in certain cell lines.

4. Replace the dye working solution with HHBS or buffer of your choice (containing an anion transporter inhibitor, such as 1 mM probenecid, if applicable) to remove any excess probes.
5. Add the stimulant as desired and simultaneously measure fluorescence using either a fluorescence microscope equipped with a FITC filter set or a fluorescence plate reader containing a programmable liquid handling system such as an FDSS, FLIPR, or FlexStation, at Ex/Em = 490/525 nm cutoff 515 nm.

### PREPARATION OF STOCK SOLUTIONS

Unless otherwise noted, all unused stock solutions should be divided into single-use aliquots and stored at -20 °C after preparation. Avoid repeated freeze-thaw cycles

#### Mag-Fluo-4 AM Stock Solution

1. Prepare a 2 to 5 mM stock solution of Mag-Fluo-4 AM in high-quality, anhydrous DMSO.

### PREPARATION OF WORKING SOLUTION

#### Mag-Fluo-4 AM Working Solution

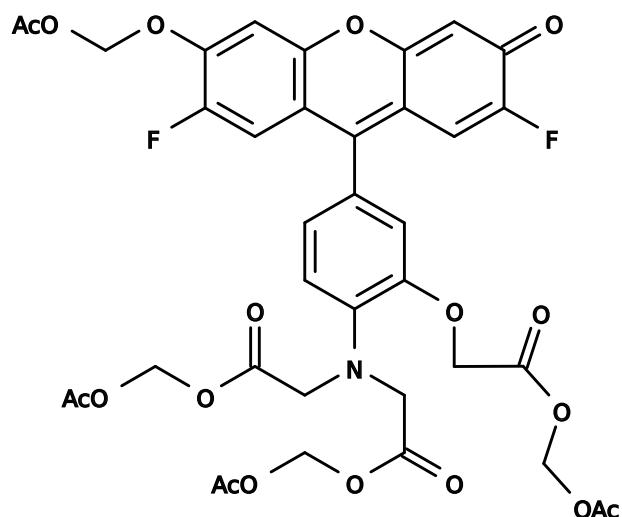
1. On the day of the experiment, either dissolve Mag-Fluo-4 AM in DMSO or thaw an aliquot of the indicator stock solution to room temperature.
2. Prepare a 2 to 20  $\mu$ M Mag-Fluo-4 AM working solution in a buffer of your choice (e.g., Hanks and Hepes buffer) with 0.04% Pluronic® F-127. For most cell lines, Mag-Fluo-4 AM at a final concentration of 4-5  $\mu$ M is recommended. The exact concentration of indicators required for cell loading must be determined empirically.

**Note:** The nonionic detergent Pluronic® F-127 is sometimes used

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## EXAMPLE DATA ANALYSIS AND FIGURES



**Figure 1.** Chemical structure for Mag-Fluo-4 AM

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