

**Rhod-2, AM \*UltraPure Grade\* \*CAS#: 145037-81-6\***

Catalog number: 21062, 21063, 21064  
Unit size: 1 mg, 50 mg, 20x50 ug

Component	Storage	Amount (Cat No. 21062)	Amount (Cat No. 21063)	Amount (Cat No. 21064)
Rhod-2, AM	Freeze (< -15 °C). Minimize light exposure	1 vial (1 mg)	1 vial (50 mg)	20x50 ug

## OVERVIEW

Calcium measurement is critical for numerous biological investigations. Fluorescent probes that show spectral responses upon binding  $\text{Ca}^{2+}$  have enabled researchers to investigate changes in intracellular free  $\text{Ca}^{2+}$  concentrations by using fluorescence microscopy, flow cytometry, fluorescence spectroscopy and fluorescence microplate readers. The long-wavelength Rhod-2  $\text{Ca}^{2+}$  indicators are valuable alternatives to Fluo-3 for experiments in cells and tissues that have high levels of autofluorescence. Rhod-2 AM is cell-permeable version of Rhod-2.

## KEY PARAMETERS

### Fluorescence microscope

Emission	TRITC filter set
Excitation	TRITC filter set
Recommended plate	Black wall/clear bottom

**Fluorescence microplate reader**

Cutoff	570
Emission	590
Excitation	540
Recommended plate	Black wall/clear bottom
Instrument specification(s)	Bottom read mode/Programmable liquid handling

## PREPARATION OF STOCK SOLUTIONS

*Unless otherwise noted, all unused stock solutions should be divided into single-use aliquots and stored at -20 °C after preparation. Avoid repeated freeze-thaw cycles*

### Rhod-2 AM \*UltraPure Grade\* Stock Solution

1. Prepare a 2 to 5 mM stock solution of Rhod-2 AM in high-quality, anhydrous DMSO.

### PREPARATION OF WORKING SOLUTION

### Rhod-2 AM \*UltraPure Grade\* Working Solution

1. On the day of the experiment, either dissolve Rhod-2 AM in DMSO or thaw an aliquot of the indicator stock solution to room temperature.
2. Prepare a 2 to 20  $\mu\text{M}$  Rhod-2 AM working solution in a buffer of your choice (e.g., Hanks and Hepes buffer) with 0.04% Pluronic® F-127. For most cell lines, Rhod-2 AM at a final concentration of 4-5  $\mu\text{M}$  is recommended. The exact concentration of indicators required for cell loading must be determined empirically.

**Note:** The nonionic detergent Pluronic® F-127 is sometimes used to increase the aqueous solubility of Rhod-2 AM. A variety of [Pluronic® F-127 solutions](#) can be purchased from AAT Bioquest.

**Note:** If your cells contain organic anion-transporters, probenecid (1-2 mM) may be added to the dye working solution (final in well

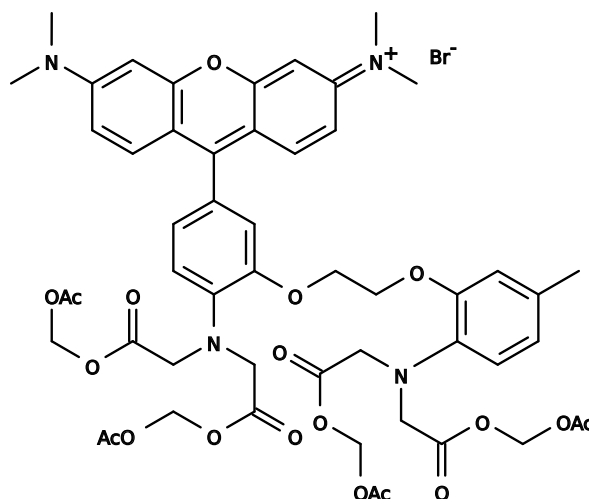
concentration will be 0.5-1 mM) to reduce leakage of the de-esterified indicators. A variety of [ReadiUse™ Probenecid products](#), including water-soluble, sodium salt, and stabilized solutions, can be purchased from AAT Bioquest.

## SAMPLE EXPERIMENTAL PROTOCOL

Following is our recommended protocol for loading AM esters into live cells. This protocol only provides a guideline and should be modified according to your specific needs.

1. Prepare cells in growth medium overnight.
2. On the next day, add 1X Rhod-2 AM working solution to your cell plate.  
**Note:** If your compound(s) interfere with the serum, replace the growth medium with fresh HHBS buffer before dye-loading.
3. Incubate the dye-loaded plate in a cell incubator at 37 °C for 30 to 60 minutes.  
**Note:** Incubating the dye for longer than 1 hour can improve signal intensities in certain cell lines.
4. Replace the dye working solution with HHBS or buffer of your choice (containing an anion transporter inhibitor, such as 1 mM probenecid, if applicable) to remove any excess probes.
5. Add the stimulant as desired and simultaneously measure fluorescence using either a fluorescence microscope equipped with a TRITC filter set or a fluorescence plate reader containing a programmable liquid handling system such as an FDSS, FLIPR, or FlexStation. at Ex/Em = 540/590 nm cutoff 570 nm.

## EXAMPLE DATA ANALYSIS AND FIGURES



**Figure 1.** Chemical structure for Rhod-2, AM \*UltraPure Grade\* \*CAS#: 145037-81-6\*

**DISCLAIMER**

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