Screen Quest™ Live Cell Chemokine (CC) receptor CCR1 cAMP Assay Service Pack

Catalog number: 38200
Unit size: 100 Tests

Component	Storage	Amount (Cat No. 38200)
Component A: Gα16 DNA	Freeze (< -15 °C), Minimize light exposure	1 vial (15 µg)
Component B: Transfectamine™ 5000	Freeze (< -15 °C), Minimize light exposure	1 vial (75 µL)
Component C: Calbryte™ 520 NW	Freeze (< -15 °C), Minimize light exposure	1 vial
Component D: 10X Pluronic® F127 Plus	Freeze (< -15 °C), Minimize light exposure	1 bottle (1 mL)
Component E: HHBS	Freeze (< -15 °C), Minimize light exposure	1 bottle (9 mL)
Component F: DMSO	Refrigerated (2-8 °C)	1 vial (100 µL)
Component G: CCR1 DNA	Freeze (< -15 °C), Minimize light exposure	1 vial (15 µg)

OVERVIEW

Chemokine (CC) receptor type 1 (CCR1), a key member of the GPCR family, is implicated in various inflammatory and immune-related disorders, making it an important therapeutic target. Traditional methods for studying GPCRs often fail to address the signaling mechanisms of specific GPCRs like CCR1, which couple to adenylyl cyclase activity and intracellular cAMP production.

The Screen Quest™ Live Cell Chemokine (CC) Receptor CCR1 cAMP Assay Service Pack is specifically designed for real-time, high-throughput monitoring of intracellular cAMP changes associated with CCR1 activation using transfected cell lines and Calbryte-520 wash-free calcium fluorescence detection methods. Unlike the conventional assays that require cell lysis, this assay preserves cellular integrity, enabling both temporal and spatial resolution of specific signaling events associated with CCR1. This assay employs cell lines that are transfected to express CCR1 along with a promiscuous G-protein Gα16. The Gα16 protein allows CCR1, which primarily signals through the cAMP pathway, to also couple to Gq signal transduction and mobilize intracellular calcium. Activation of CCR1 by specific ligands, such as CCL3 (MIP-1α) or CCL5 (RANTES), can be detected using calcium-sensitive dyes like Calbryte™ 520 AM, Cal-520™ AM, Fluo-8™ AM, Fluo-4™ AM, or corresponding no-wash calcium kits. The inclusion of CCR1 and Gα16 co-expression ensures robust calcium signaling for reliable assay performance.

The service pack provides all necessary components for precise measurement of CCR1-mediated cAMP changes using FLIPR, FDSS, or equivalent fluorescence microplate readers. It can be utilized for studying non-Gq protein-coupled CCR1 activity, offering researchers a reliable tool to explore CCR1 signaling pathways and assess potential therapeutic compounds targeting this receptor.

AT A GLANCE

Protocol summary

1. Prepare cells for transfection
2. Prepare Transfectamine™ 5000-DNA mixture
3. Add Transfectamine™ 5000-DNA mixture to the cell culture, incubate overnight
4. Transfer the transfected cells to a 96-well plate 24-30 hours after transfection, and incubate the culture overnight
5. Add Calbryte™ 520 NW dye-loading solution
6. Incubate at room temperature or 37 °C for 30-60 minutes
7. Monitor the fluorescence intensity at Ex/Em = 490/525 nm

Important Note

Thaw the kit components at room temperature before starting the experiment.

KEY PARAMETERS

Fluorescence microplate reader

Cutoff	515 nm
Emission	525 nm
Excitation	490 nm
Recommended plate	Black wall/Clear bottom
Instrument specification(s)	Bottom read mode/Programmable liquid handling

Note: This kit is compatible with the following instruments: FDSS, FLIPR, ViewLux, NOVOStar, Array Scan, FlexStation, and IN Cell Analyzer.

CELL PREPARATION

1. Seed the cells such that they will be ~60-70% confluent at the time of transfection.
2. Replace with fresh growth medium before transfection. For example, replace with 2 mL of medium per well for 6-well plates and 6 mL of medium for 10 cm plates.

PREPARATION OF STOCK SOLUTIONS

Calbryte™ 520NW stock solution

1. Add 20 µL of DMSO (Component F) into the vial of Calbryte™ 520NW (Component C), and mix them well.

   Note: 20 µL of Calbryte™ 520NW stock solution is enough for one plate. Unused Calbryte™ 520NW stock solution can be aliquoted and stored at ≤ -20 °C for more than one month, provided the tubes are tightly sealed. Protect from light and avoid repeated freeze-thaw cycles.

PREPARATION OF WORKING SOLUTION

1X Assay Buffer

1. Combine 9 mL of HHBS (Component E) with 1 mL of 10X Pluronic® F127 Plus (Component D), and mix thoroughly.

Calbryte™ 520NW Working Solution

1. Add 20 µL of Calbryte™ 520 NW stock solution to 10 mL of 1X

Assay Buffer, and mix well.

Note: The working solution is stable for at least 2 hours at room temperature.

TRANSFECTAMINE™ 5000-DNA MIXTURE

1. Add 15 µL of ddH_2O to the vial of Ga16 DNA (Component A) and CCR1 DNA (Component G), to get the final concentration of 1 µg/µL for both DNAs.
2. Mix 3 µg of DNA [for example, 1.5 µg of Ga16 DNA (Component A) and 1.5 µg of CCR1 DNA (Component G)] with 200 µL of serum-free medium.
3. Add 9 µL of Transfectamine™ 5000 (Component B) to the mixture from Step 2.
4. Mix well and incubate at room temperature for 20 minutes.

   Note: The ratio of Transfectamine™ 5000 and DNA need to be optimized for different cell lines. In general, the ratio for Transfectamine™ 5000 Transfection Reagent (µL) to DNA (µg) should be 3-5 µL : 1 µg.

Table 1. Sample protocols for a 6-well plate and a 10 cm plate

Component	6 well plate (per well)	10 cm plate
Fresh culture medium	2 mL	6 mL
Plasmid	~3 µg	10~15 µg
Serum-free medium	200 µL	600 µL
Transfectamine™ 5000 Transfection Reagent	~9 µL	~30-45 µL

SAMPLE EXPERIMENTAL PROTOCOL

Transfection protocol

1. Add Transfectamine™ 5000 -DNA mixture to the culture plate and incubate overnight.

   Note: The recombinant protein can start to be detected as early as 16 hours after transfection. The maximal expression level may be observed 72~96 hours after transfection.

2. Transfer the transfected cells to a 96-well plate 24-30 hours post transfection and incubate overnight.
   - **For adherent cells:** Plate cells overnight in the growth medium at 40,000 to 80,000 cells/well/100 µL for a 96-well plate.
   - **For non-adherent cells:** Centrifuge the cells from the culture medium and then suspend the cell pellet in cell growth medium or HHBS at 125,000 to 250,000 cells/well/100 µL for a 96-well poly-D lysine plate. Centrifuge the plate at 800 rpm for 2 minutes with brake off prior to the experiments.

   Note: Each cell line should be evaluated on the individual basis to determine the optimal cell density for the intracellular calcium mobilization.

Calcium assay

1. Add 100 µL/well (96-well plate) of Calbryte™ 520NW working solution into the cell plate.
2. Incubate the dye-loaded plate in a cell incubator for 30 minutes, and then incubate the plate at room temperature for another 15-30 minutes.

   Note: If the assay requires 37 °C, perform the experiment immediately without further room temperature incubation.

   Note: If the cells can function well at room temperature for longer time, incubate the cell plate at room temperature for 1 hour (It is recommended that the incubation time be no longer than 2 hours.)

3. Prepare the compound plate with HHBS or your desired buffer.
4. Run the calcium flux assay by monitoring the fluorescence intensity at Ex/Em = 490/525 nm.

EXAMPLE DATA ANALYSIS AND FIGURES

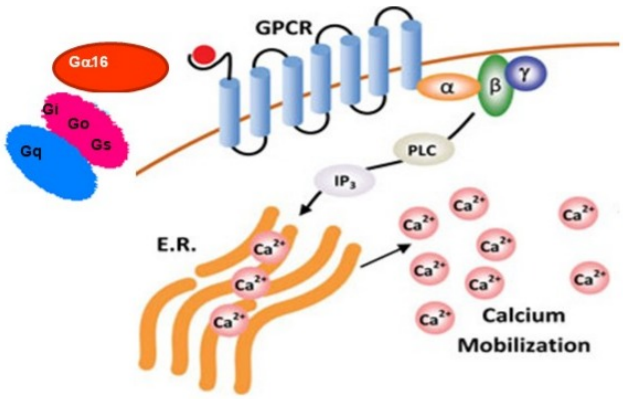

Figure 1. Screen Quest™ Live Cell Chemokine (CC) receptor CCR1 cAMP assay principle

DISCLAIMER

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